Frequent EPAS1/HIF2A exon 9 and 12 mutations in non-familial pheochromocytoma

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Abstract

Pheochromocytomas are neuroendocrine tumors arising from the adrenal medulla. While heritable mutations are frequently described, less is known about the genetics of sporadic pheochromocytoma. Mutations in genes involved in the cellular hypoxia response have been identified in the tumors, and recently EPAS1, encoding HIF-2α, was revealed as a new gene involved in the pathogenesis of pheochromocytoma and abdominal paraganglioma. The aim of this study was to further characterize EPAS1 alterations in non-familial pheochromocytomas. Tumor DNA from 42 adrenal pheochromocytoma cases with apparently sporadic presentation, without known hereditary mutations in predisposing genes, were analyzed for mutations in EPAS1 by sequencing of exons 9 and 12, which contain the two hydroxylation sites involved in HIF-2α degradation, and also exon 2. In addition, copy number at the EPAS1 locus as well as transcriptome-wide gene expression was studied by DNA and RNA microarray analysis, respectively. We identified six missense EPAS1 mutations, three in exon 9 and three in exon 12, in five of 42 pheochromocytomas (12%). The mutations were both somatic and constitutional, and had no overlap to 11 cases (26%) with somatic mutations in NF1 or RET. One sample had two different EPAS1 mutations, shown by cloning to occur in cis, possibly suggesting a novel mechanism of HIF-2α stabilization through inactivation of both hydroxylation sites. One of the tumors with EPAS1 mutation also had DNA copy number gain at the EPAS1 locus. All EPAS1-mutated tumors displayed a pseudo-hypoxic gene expression pattern, suggestive of an oncogenic role of the identified mutations.
Introduction

Pheochromocytomas and abdominal paragangliomas are catecholamine-producing tumors derived from neuroendocrine chromaffin cells in the adrenal medulla or the extra-adrenal paraganglia. The tumors can be a manifestation of different hereditary tumor syndromes such as multiple endocrine neoplasia type 2 (MEN2), von Hippel-Lindau disease (VHL), neurofibromatosis type 1 (NF1) or familial pheochromocytoma-paraganglioma syndrome, which are caused by mutations in the genes RET, VHL, NF1 and SDHx, respectively (Dahia 2014; Welander et al. 2011). In recent years, additional susceptibility genes for pheochromocytoma and paraganglioma have been discovered, including TMEM127 (Qin et al. 2010), MAX (Comino-Mendez et al. 2011) and FH (Castro-Vega et al. 2014), and more rarely EGLN1 (Ladroue et al. 2008) and KIF1Bβ (Schlisio et al. 2008). The majority of pheochromocytomas and paragangliomas are apparently sporadic, i.e. isolated and non-familial. Until recently, somatic mutations in many of the genes known to be associated with familial disease were only found in a small proportion of the cases (Burnichon et al. 2011). This changed when somatic NF1 mutations were revealed in about 20-25% of sporadic pheochromocytomas (Burnichon et al. 2012; Welander et al. 2012), but still the majority of sporadic tumors remain unexplained. Studies of gene expression have revealed that pheochromocytomas and paragangliomas can be divided into two groups: VHL- and SDHx-mutated tumors form one cluster (Cluster 1) with a hypoxia-related transcription signature, whereas RET/NF1/TMEM127/MAX-mutated tumors form another cluster (Cluster 2) and display transcription profiles characterized by the activation of kinase signalling pathways (Dahia 2014; Dahia et al. 2005; Eisenhofer et al. 2004). Sporadic tumors cluster into either of the two distinct groups instead of forming clusters of their own, indicating that similar molecular mechanisms are involved.
Hypoxia-inducible factors (HIFs) are transcription factors involved in the cellular hypoxia response. Active HIF is a dimer composed of an α and a β subunit. Whereas the β subunit is stably expressed, the levels of the α subunits HIF-1α, HIF-2α and HIF-3α are regulated by oxygen-dependent prolyl hydroxylases (Kaelin and Ratcliffe 2008; Keith et al. 2012). Prolyl hydroxylation occurs at two specific proline residues in the oxygen-dependent degradation domain of HIF-α. This allows recognition and ubiquitination of the protein by the VHL tumor suppressor E3 ubiquitin ligase complex, thereby targeting it for proteasomal degradation. At low oxygen levels the prolyl hydroxylation is reduced, resulting in stabilization of HIF-α and subsequent transcription of target genes involved in the hypoxia response, including metabolic changes and angiogenesis.

Mutations in VHL, EGLN1 (encoding the prolyl hydroxylase PHD2) and SDHx (encoding the subunits of succinate dehydrogenase) all result in an accumulation of HIF-α and thus a pseudo-hypoxic response, allowing activation of target genes hypothesized to contribute to chromaffin cell tumorigenesis (Maher 2013). Rare patients with polycythemia, a disease state characterized by an elevated concentration of red blood cells, have mutations in VHL, EGLN1 or in EPAS1, the gene encoding HIF-2α (Bento et al. 2014). Recently, somatic gain-of-function mutations in EPAS1 were reported for the first time in paragangliomas associated with polycythemia (Zhuang et al. 2012), providing an additional link between polycythemia and neuroendocrine tumors and also supporting the hypothesis of pseudo-hypoxia as an important process in their development. A subsequent study revealed a germline EPAS1 mutation in a patient with polycythemia and paraganglioma (Lorenzo et al. 2013) which, just like the first mutations identified, was demonstrated to decrease ubiquitination and degradation, thereby increasing the stability of HIF-2α. Subsequent studies identified somatic
EPAS1 mutations in pheochromocytomas and paragangliomas also in the absence of polycythemia (Comino-Mendez et al. 2013; Favier et al. 2012; Toledo et al. 2013), and demonstrated that EPAS1 mutations promote tumor growth in mice (Toledo et al. 2013). So far, only a few cases of EPAS1 mutations have been reported in single sporadic pheochromocytomas/paragangliomas without polycythemia. Here, we further investigated this new class of mutations by analyzing EPAS1 in a cohort of adrenal pheochromocytomas from patients with isolated, non-familial disease without polycythemia. In contrast to several previous studies focusing on exon 12 only, we sequenced both exons that cover the HIF-2α hydroxylation sites, as a gain-of-function mutation has once been described in exon 9 (Lorenzo et al. 2013). As one mutation of unknown pathogenicity has been described in exon 2, adjacent to the DNA binding domain (Toledo et al. 2013), this exon was also included. The findings were evaluated in view of copy number and gene expression results, clinical presentation, and previously detected somatic mutations in these tumors.

Materials and Methods

Patients and biological samples

The study included 42 primary adrenal pheochromocytomas from 42 patients presenting as sporadic without family history or syndromic features of the disease. None of the patients had polycythemia. Clinical data for the cohort has previously been published (Welander et al. 2012) and is presented in detail in Supplementary Table S1. Two patients had developed metastases during follow-up and were therefore classified as malignant according to the WHO criteria (DeLellis et al. 2004), while 40 cases were without metastases. In addition, all cases were classified according to AFIP criteria (Lack 2007) (Supplementary Table S1). All samples were collected and studied with informed consent and approval from the local ethic
committees. DNA was extracted from tissue and blood samples using the Maxwell 16 Tissue DNA Purification Kit (Promega, Madison, WI) and the Maxwell 16 Blood DNA Purification Kit (Promega), respectively. RNA was extracted using the RNeasy Minikit (Qiagen, Hilden, Germany), and cDNA synthesis was performed with the Maxima First Strand cDNA synthesis kit (Thermo Scientific, Waltham, MA).

The tumors have previously been investigated for mutations in the genes NF1, RET, VHL, SDHB, SDHD and MAX, which revealed somatic NF1 mutations in 10 samples and a somatic RET mutation in one sample (Welander et al. 2012). Two patients had non-synonymous single nucleotide polymorphisms in SDHB (rs11203289) and SDHD (rs11214077), respectively. Subsequent mutation analysis of TMEM127 in all cases revealed a novel, constitutional missense sequence variant in one sample (c.10C>T, Supplementary Figure S1), which was predicted as benign by the PolyPhen-2 algorithm (Adzhubei et al. 2010).

**Mutation screening and DNA copy number analysis of EPAS1**

Exons 2, 9 and 12 of EPAS1 were analyzed by direct Sanger sequencing of tumor DNA using previously described methodology (Welander et al. 2012) and primers as specified in Supplementary Table S2. Mutations were confirmed by sequencing of a second, independent PCR product from the same sample. When available, mutation analysis was also performed in corresponding normal DNA from patients with mutations in the tumor DNA. In case 38, which had two different mutations, PCR amplification and cloning of EPAS1 cDNA into *Escherichia coli* was performed to determine if the mutations occurred in cis or trans. A previously described cloning technique (Welander et al. 2012) was used, with the primers 5’-CGTCCTGAGTGAGATTGAGAAG-3’ (in the exon 8-9 boundary) and 5’-
CCAGTGCTGGAAGATGTTT -3’ (in exon 12). *EPAS1* cDNA was then PCR amplified from each clone and sequenced.

The effects of missense mutations were predicted with PolyPhen-2 (Adzhubei et al. 2010). To study conservation of the affected residues, a multiple sequence alignment of the HIF-2α amino acid sequence between different species and a comparison with the human HIF-1α protein was performed using the Clustal Omega algorithm (Sievers et al. 2011). Sequences used were HIF-2α (EPAS1) from *Homo sapiens* (UniProt accession number Q99814), *Mus musculus* (P97481), *Bos Taurus* (A5PJ71) *Gallus gallus* (Q9W7C6) and *Danio rerio* (B3DJ1), together with HIF-1α from *Homo sapiens* (Q16665) for comparison.

Previously retrieved data from SNP microarray analysis (Affymetrix GeneChip Human Mapping 250K) of tumor DNA (Welander et al. 2012) was analyzed for copy number alterations in *EPAS1* using Genotyping Console v. 4.0 (Affymetrix, Santa Clara, CA).

**Gene expression analysis**

Total RNA was quantified with a NanoDrop-1000 spectrophotometer (Thermo Scientific) and 250 ng were used for whole-transcriptome analysis with GeneChip Human Gene 1.0 ST arrays (Affymetrix), covering 28,869 annotated genes with a median of 26 probes per gene.

Two of the 42 sporadic tumor samples (number 7 and 20) for which no RNA was available were excluded. Five hereditary tumors were used as controls (Supplementary Table S3): two with RET mutations, one with *NF1* mutation (Welander et al. 2012), one with *VHL* mutation and one with *SDHA* mutation (Welander et al. 2013). Included samples had RNA integrity numbers (RIN) between 6.6 and 9.1 (mean 8.3) when analyzed with a 2100 Bioanalyzer system using the Eukaryote Total RNA Nano assay (Agilent Technologies, Santa Clara, CA).
Sense-strand cDNA was generated using the Ambion WT Expression Kit (Life Technologies, Carlsbad, CA) and then fragmented, labelled and hybridized to arrays in a GeneChip Hybridization Oven 640 using the GeneChip WT Terminal Labeling Kit (Affymetrix), all according to the protocols supplied by the manufacturers. The arrays were washed and stained in a GeneChip Fluidics Station 450 and scanned in a GeneChip Scanner 3000 7G (Affymetrix).

Microarray data files (CEL) were opened in Expression Console v. 1.3 (Affymetrix) and normalized using the robust multi-array average (RMA) algorithm. Normalized signal intensities for EPAS1, its target genes VEGFA, CCND1, EDN1, EPO and SLC2A1 (Jochmanova et al. 2013), and the gene for phenylethanolamine N-methyltransferase (PNMT) were exported for separate statistical analysis as described below. The CEL files were also imported into GeneSpring GX v. 12.6 (Agilent Technologies) for transcriptome-wide statistical analysis and clustering. After RMA normalization, a quality filter was applied to remove probesets for which none of the 45 samples had signal intensity values greater than the 20th percentile of all signal intensity values of the sample. To test how the transcription profiles of EPAS1-mutated samples would cluster compared to genotypes with previously known expression patterns, a set of genes previously shown to separate the SDHx, VHL and RET/NFL groups were downloaded (Burnichon et al. 2011). These genes overlapped with 454 probesets that had passed the quality filter in our analysis, which were used to perform a hierarchical clustering with a Euclidean distance metric and Ward’s linkage rule.

Statistical analysis

Gene expression levels of the seven candidate genes described above were compared between EPAS1-mutated and EPAS1-wildtype sporadic tumors using two-tailed Student’s t-tests on the
exported log2 signal values. After back-transformation ($2^{(\log_2 \text{signal})}$), expression levels were normalized to the mean of the three hereditary Cluster 2 controls (samples 8, 12 and 14) for visualization in GraphPad Prism v.6.03. Transcriptome-wide statistical analysis was performed on all genes passing the quality filter in GeneSpring, where genes with differential expression between $EPAS1$-mutated and $EPAS1$-wildtype sporadic tumors were identified by $t$-tests. The Benjamini-Hochberg method (Benjamini and Hochberg 1995) was used to control for multiple testing. Genes were regarded as differentially expressed when having a false discovery rate (FDR) <0.05 and an absolute fold change >2.

The clinical variables age, tumor size and hormone levels were compared between cases with and without $EPAS1$ mutations using two-tailed Student’s $t$-tests. Fisher’s exact test was used to test for association with malignancy and gender. P-values <0.05 were regarded as significant.

**Results**

**Mutations in $EPAS1$**

Among 42 non-familial pheochromocytomas from 42 unrelated patients, five tumors harboured six missense mutations in $EPAS1$ (Table 1, Fig. 1A), of which three were found in exon 9 and three in exon 12. No mutations were detected in exon 2. Two of the mutations were present in blood or normal tissue whereas two were confirmed somatic, and two are unknown because neither blood nor non-tumor tissue was available (Fig. 1B). Two of the $EPAS1$ mutations occurred in the same patient, of which one was present in DNA from blood whereas the other was somatic. Molecular cloning showed that the two mutations occurred in $cis$ (Supplementary Figure S2). All but one of the identified mutations occurred at highly
conserved residues, and the three mutations in exon 12 were all in or in the vicinity of the primary hydroxylation site (Fig. 1C). PolyPhen analysis predicted four of the six amino acid substitutions as damaging (Table 1).

Based on SNP microarray data, case 11, which also carried a mutation, had gain (three copies) of the 2p21 chromosomal region where EPAS1 is located (Supplementary Figure S3). The other 41 samples showed no copy number gain, but two samples without mutations (cases 2 and 34) had heterozygous loss of 2p, including EPAS1. No copy number alterations of the EPAS1 locus were found in 60 blood DNA samples from healthy controls analyzed by SNP microarrays. Previously known intronic or synonymous polymorphisms in EPAS1 (rs7557402, rs116510029, rs35606117, rs184760160 and rs41281469) were detected in expected frequencies according to information in the Ensembl database.

Gene expression patterns in EPAS1-mutated pheochromocytomas

Tumors with EPAS1 mutations had a significantly higher gene expression of both EPAS1 (P=0.0031) and its target gene VEGFA (P=0.0039) than tumors without mutations (Figure 2A). In addition, the EPAS1-mutated tumors had a very low expression of PNMT (P<0.0001). Of note, the target genes CCND1, EDN1, EPO and SLC2A1 (Jochmanova et al. 2013) which have been frequently investigated by others (Lorenzo et al. 2013; Toledo et al. 2013; Zhuang et al. 2012), did not show any difference in expression in this study (Supplementary Figure S4). Possible explanations may include differences in detection methods or sampling, and/or that these genes, though clearly regulated by HIF-2α in some tissues, may have a more variable expression in the adrenal medulla. Six tumors without identified mutations but with high EPAS1 expression were sequenced for all the remaining exons of EPAS1 (Supplementary Table S2). No mutations were detected, but one case (case 6) had a missense
polymorphism in exon 15 predicted as benign (rs59901247, PolyPhen score 0.001, allele frequency 0.15 in Ensembl).

One of the recent large gene expression studies of pheochromocytomas and paragangliomas identified a set of genes that could separate tumors with SDHx, VHL and RET/NF1 mutations by hierarchical gene expression clustering (Burnichon et al. 2011). To investigate how the gene expression profiles of EPAS1-mutated tumors would cluster compared to the previously known genotypes, we utilized the same set of genes (454 probesets) for hierarchical clustering of the present cohort. As expected, tumors with known somatic NF1 and RET mutations clustered together with hereditary NF1 and MEN2A cases (Figure 2B). In contrast, all five tumors with EPAS1 mutations clustered together with the hereditary VHL and SDH cases, in agreement with the hypothesis of a pseudo-hypoxic behaviour. The sporadic tumors without identified mutations were distributed between both the main clusters. After transcriptome-wide comparison of sporadic tumors with and without EPAS1 mutations, 22 genes were significantly and more than 2-fold differentially expressed between the groups (Supplementary Table S4) after correction for multiple testing. Among those upregulated in EPAS1 cases were several genes involved in cell metabolism (COX4I2, NOX4, FOLH1, FOLH1B, COX17) and angiogenesis (KDR/VEGFR, ANGPT2). The most significantly upregulated gene, COX4I2, has previously been reported to be strongly upregulated in VHL/SDH-related pheochromocytomas (Favier et al. 2009) and another one, GNA14, was recently observed by others to be significantly upregulated in EPAS1-mutated tumors (Comino-Mendez et al. 2013). Among the most downregulated genes are NCAM2 (a neural cell adhesion molecule, previously found upregulated in Cluster 2 tumors (Burnichon et al. 2011)), HCN1 (a cation channel contributing to neuron currents), and PNMT.
Clinical presentation of EPAS1-mutated pheochromocytomas

All five patients with EPAS1 mutation(s) had a single pheochromocytoma without recurrence or metastasis, and none suffered from polycythemia. Re-evaluation of medical records showed that none of the patients had signs of hereditary tumor disease, but one presented with a ductal breast cancer in situ four years after the pheochromocytoma diagnosis (patient 22) and one suffered from sickle cell anemia (patient 52). Histopathologically, it can be noted that the tumor from case 19 showed slightly increased proliferation with 5% Ki-67 positive cells, and case 22 presented local invasion in agreement with histopathological malignancy according to AFIP criteria (Lack 2007). Patients with EPAS1 mutations tended to be younger than those without EPAS1 mutations (mean 52.2±7.2 years vs. 61.6±12.9 years) but the difference was not statistically significant (P=0.12). No associations were found between EPAS1 mutation status and tumor size, malignancy or gender, but it can be noted that 80% of the patients with EPAS1 mutations were females, in agreement with the female dominance in the cases reported so far (Comino-Mendez et al. 2013; Pacak et al. 2013; Toledo et al. 2013).

However, in another cohort where all susceptibility genes were investigated, EPAS1 mutations were detected in four males and one female (Welander et al. 2014). Data on hormone profiles was incomplete and did not allow any statistically significant conclusions. It can be noted that all five patients with EPAS1 mutations were observed with increased norepinephrine levels (Table 1, Supplementary Table S1). In addition, these patients had normal or only slightly increased epinephrine levels (Table 1, Supplementary Table S1), which is in consistence with their low expression of PNMT (Fig. 2A).

None of the cases with EPAS1 mutation(s) had mutations in any of the tested susceptibility genes NF1, RET, VHL, SDHB, SDHD or MAX, i.e. they were mutually exclusive with the somatic mutations identified in NF1 (24%) and RET (2.4%) in the cohort (Supplementary
Table S1 and Figure S5). One case (number 19) had a constitutional, probably benign
sequence variant in \textit{TMEM127} (Supplementary Figure S1).

\textbf{Discussion}

Mutation analysis of \textit{EPAS1} in 42 non-familial pheochromocytomas resulted in identification
of six different mutations, of which three have not been reported earlier in
pheochromocytoma or paraganglioma. The three mutations identified in exon 12 occur in or
in close proximity to the primary hydroxylation site. Thus, they are likely to affect the
conformation of the hydroxylation domain and disrupt prolyl hydroxylation and degradation
of HIF-2α, as has previously been functionally demonstrated for this type of mutations
was predicted as non-pathogenic by PolyPhen and has been reported as a very rare
polymorphism (rs150797491 with allele frequency 0.00092 in Ensembl), but, nevertheless, it
has been demonstrated as a gain-of-function mutation that decreases VHL protein binding and
ubiquitination, increasing the stability of HIF-2α (Lorenzo \textit{et al.} 2013). In both this and the
previous study, the mutation was present in the constitutional DNA of a
pheochromocytoma/paraganglioma patient. Met368Ile has also been reported as a rare
polymorphism (rs61757375 with allele frequency 0.0014). This could be in agreement with
the proposition that \textit{EPAS1} mutations are pheochromocytoma/paraganglioma-promoting, but
not sufficient for tumor development (Lorenzo \textit{et al.} 2013), but further studies will be
required to determine its pathogenicity. The novel Ile412Asn mutation occurs in a highly
conserved residue close to the hydroxylation site and is bioinformatically predicted to be
damaging.
Interestingly, two heterozygous EPAS1 mutations were identified in cis in the same tumor. It seems probable that the constitutional mutation, affecting an amino acid seven residues from the Pro405 hydroxylation site, is less severe than the somatic mutation close to the primary hydroxylation site Pro531 (Fig. 1C). Speculatively, the combination of the two may lead to an even higher stability of the HIF-2α protein than one mutation would cause alone, as previous results have suggested for the HIF-1α protein (Chan et al. 2005). One patient with an EPAS1 mutation also had copy number gain of the EPAS1 locus. The combination of activating mutations and copy number gain has previously been reported for EPAS1 (Comino-Mendez et al. 2013), as well as for other oncogenes (Modrek et al. 2009).

Our findings are in agreement with previous reports (Comino-Mendez et al. 2013; Toledo et al. 2013), and merging the present study with the two previous ones implies a total frequency of EPAS1 mutations of around 5.3% in sporadic tumors. In this cohort, the frequency of EPAS1 mutations was 11.9% or, more conservatively, 7.1% when not considering the two variants not immediately in the hydroxylation sites for which no functional studies have been performed. EPAS1 mutations were mutually exclusive with somatic mutations in other susceptibility genes in our cohort of pheochromocytomas, supporting an oncogenic role.

All tumors with EPAS1 mutations showed an increased gene expression of the HIF-2α target gene VEGFA, which has also previously been shown to be upregulated in pheochromocytomas and paragangliomas with EPAS1 mutations (Lorenzo et al. 2013; Toledo et al. 2013; Zhuang et al. 2012). Notably, the tumors with mutations also had a higher expression of EPAS1 itself. The reason for this is not clear, as the mutations are thought to act on the protein level, but the same result has been observed previously (Comino-Mendez et al. 2013). We also show that the tumors with EPAS1 mutations have a very low expression of
PNMT, encoding the enzyme phenylethanolamine N-methyltransferase which catalyzes the methylation of norepinephrine to form epinephrine. This is in agreement with the loss of PNMT expression in other Cluster 1 tumors as compared to Cluster 2 tumors which have a high expression, resulting in high epinephrine levels (Eisenhofer et al. 2004). Further, the five tumors with EPAS1 mutations clustered together with hereditary VHL and SDH cases, supporting that the mutations have a role in the disease by inducing a similar pseudo-hypoxic response. Interestingly, some sporadic tumors without any known mutations also clustered with the VHL/SDH/EPAS1 cases, some of which also had an increased expression of EPAS1. Those with high EPAS1 expression were sequenced for all EPAS1 exons, but no additional mutations were identified. This suggests that other, so far unknown, alterations of hypoxia response mechanisms may be involved in a subset of the sporadic tumors.

EPAS1 mutations have now been identified in association with a variety of phenotypes, including polycythemia without tumors (Percy et al. 2008), polycythemia in combination with single or multiple pheochromocytomas/paragangliomas and sometimes multiple somatostatinomas (Pacak et al. 2013; Taieb et al. 2013; Yang et al. 2013; Zhuang et al. 2012), as well as single or multiple pheochromocytomas/paragangliomas without polycythemia (Comino-Mendez et al. 2013; Toledo et al. 2013). Previous findings of EPAS1 mutations in different tumors from the same patient, but not in germline DNA, indicate that somatic EPAS1 mutations may occur in a cell during embryogenesis and predispose the affected tissues to tumor formation (Zhuang et al. 2012). However, it has been demonstrated that at least one reported case harboured a somatic mutation only in the tumor and not in adjacent normal tissue (Comino-Mendez et al. 2013). Possibly the variable phenotypes, constituted by different combinations of polycythemia and/or single or multiple tumors, may be directed by the specific time point of a somatic EPAS1 mutation, e.g. during early embryogenesis or later
in life. Plausibly, the type of mutation as well as additional genetic, epigenetic or environmental factors are also part of the explanation. In addition to somatic mutations, one case with an inherited mutation, identical to one of the constitutional mutations reported in this study, has been described before (Lorenzo et al. 2013), but in contrast to the study by Lorenzo et al, the patient reported here did not suffer from polycythemia. The penetrance of pheochromocytoma/paraganglioma for constitutional mutations is still unknown and will require further family studies, but the lack of family history of disease in the cases reported here support an incomplete penetrance. It can also not be excluded that some of the variants do not play a pathogenic role in the clinical phenotype.

In conclusion, we show that somatic as well as germline EPAS1 mutations are of importance in pheochromocytoma development. Our results indicate that mutations in both exon 9 and exon 12 of EPAS1 are frequent in isolated, non-familial pheochromocytomas, and that they are associated with a pseudo-hypoxia-related gene expression pattern.

Declaration of interest

The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the research reported.

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Figure legends

Figure 1. Mutations in EPAS1 identified in adrenal pheochromocytomas with sporadic presentation. A) Mutations identified in tumor DNA. B) Sequences from corresponding normal DNA (when available), showing that two of the mutations are constitutional. C) Multiple sequence alignment of HIF-2α (EPAS1) from different species together with the human HIF-1α amino acid sequence. Prolyl hydroxylation occurs at Pro405 and Pro531 in human HIF-2α, indicated with blue color. Symbols: [*], identical residues; [:], conserved substitutions; [,], semi-conserved substitutions.

Figure 2. Gene expression in pheochromocytomas in relation to EPAS1 mutation status. A) RNA levels of the genes EPAS1, VEGFA and PNMT compared between sporadic pheochromocytomas with (EPAS1 mut, n=5) and without (EPAS1 wt, n=35) EPAS1 mutations. The gene expression has been normalized to the mean value of cases with hereditary NF1 and MEN2A. Horizontal bars represent mean values. B) Hierarchical clustering of tumors based on RNA expression levels of 454 genes (implicated in pheochromocytoma subgroups (Burnichon et al. 2011)). High and low expression is indicated by green and red color, respectively. Cases with mutations are highlighted below the dendrogram.
Table 1. Details for apparently sporadic pheochromocytomas with mutation(s) in *EPAS1/HIF2A*.

<table>
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<th>Case number</th>
<th>Gender</th>
<th>Age</th>
<th>Malignancy, size</th>
<th>Exon</th>
<th>Mutation&lt;sup&gt;a&lt;/sup&gt;</th>
<th>Present in normal DNA&lt;sup&gt;b&lt;/sup&gt;</th>
<th>PolyPhen prediction (score)</th>
<th><em>EPAS1</em> gain</th>
<th>Biochemical phenotype NE / EPI / DA</th>
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</tbody>
</table>

F, Female; M, Male. EPI, epinephrine levels in plasma or urine before surgery; NE, norepinephrine levels in plasma or urine before surgery; DA, dopamine levels in urine before surgery; N, normal; I, increased, -, no value (further details can be found in Supplementary Table S1).

<sup>a</sup>Mutation nomenclature is based on the human *EPAS1* sequence ENST00000263734 in the Ensembl database.

<sup>b</sup>Non-tumor DNA was extracted from blood (samples 19 and 38) or adrenal cortex (sample 52).

<sup>c</sup>Previously reported in pheochromocytoma/paraganglioma (Comino-Mendez *et al.* 2013; Toledo *et al.* 2013).

<sup>d</sup>Previously reported in pheochromocytoma/paraganglioma (Lorenzo *et al.* 2013).

<sup>e</sup>Malignant according to AFIP criteria (Lack 2007), local invasion.

<sup>f</sup>Previously reported in pheochromocytoma/paraganglioma (Yang *et al.* 2013).