Genetic variants in thyroid cancer distant metastases

Dear Editor,

Thyroid carcinoma is the most rapidly increasing solid tumor in the United States; although there has been a great emphasis on the analysis of primary tumors, predictors of both late-stage progression and response to therapy are more poorly defined due in part to the scarcity of progressive metastatic tissues. Therapeutic evidence of mixed responses in metastatic lesions and the limited available genomic data suggest that distant metastases in thyroid cancer are heterogeneous, driven by known oncogenes and other pathways that also might be therapeutic targets. We analyzed the genomes of a small number of rare surgically resected metastatic thyroid cancer lesions along with paired normal and primary tumor samples when available, in an effort to better characterize progressive distant metastases. The findings confirm the presence of mutations to known tumor drivers (*BRAF* and *RAS*) in metastatic samples. The results also identified the co-occurrence in predicted functional variants to the DNA damage repair (DDR) genes *ATM* and *ERCC4* in metastatic lesions that did not show alterations of the MAPK pathway.

We examined the exomes of 19 samples (including 5 paired normal tissues) from 11 follicular cell-derived thyroid cancer patients with surgically resected distant metastases by custom exomeSeq (Supplementary Materials and Methods, see section on supplementary data given at the end of this article). Tumors with different histopathologies were included and were confirmed by an expert thyroid pathologist (PW). All patients were treated with TSH suppression, eight of them received I-131 therapy and none of them received chemotherapy or kinase inhibitors. Overall, 19,299 unique variants in the 682 genes and genomic regions were identified by an exomeSeq custom panel. We sought to identify rare, conserved, exonic variants that were likely to have functional effects. Thus, we focused on 1742 exonic variants in exons of sequenced genes. We excluded synonymous variants and variants with higher than 0.01 minor allelic frequency in the 1000 Genomes

Project. We then filtered to include variants located in regions that were conserved through vertebrate evolution. These criteria produced a list of 349 variants in 199 genes. Unsupervised clustering using complete linkage and Euclidian distance identified clusters based on patients, suggesting that most variants were primarily associated with individuals rather than histology or tissue location (data not shown). We finally applied a filter to identify variants predicted or known to have a damaging (SIFT) or deleterious (PolyPhen) effect on the protein. Figure 1 summarizes the filtering strategy.

In the primary tumors of patients with distant metastases, 33 variants in 31 genes were identified by comparing with normal samples. We focused on genes affected in more than one tumor sample and found three such genes: *BCR*, *BRAF* and *MAP4* (Fig. 2). Two variants that affect *BRAF* in five primary tumor samples were identified. A Hürthle cell carcinoma (HCC) bore a *BRAF T241M* variant, although its functional significance is uncertain. The remaining samples with *BRAF* had mutations resulting in *BRAF V600E* and included two papillary thyroid cancers (PTC) primary tumors and a metastatic lesion, and one anaplastic thyroid cancer (ATC) metastatic lesion. We also identified four samples from four patients with an identical insertion resulting in a frameshift (chr22:23653975 Indel: TCCGG) in *BCR* including three primary tumors and one metastasis. The primary tumor samples were from follicular (FTC) and HCC, and a metastatic lesion from *BRAF V600E* PTC. Finally, three primary tumors had an identical mutation (R1112P) in *MAP4*. Two of the tumors are from patients with HCC, and the third is a PTC with a concurrent *BRAFV600E* mutation. *BCR* and *MAP4* are functionally involved in cytoskeletal dynamics, although those roles and functional implications of the variants identified are not completely characterized.

In the metastatic tissues, we identified thirty variants in twenty-eight genes after filtering unique vs normal tissues. Variants in three genes, *ATM*, *BRAF* and *ERCC4*,
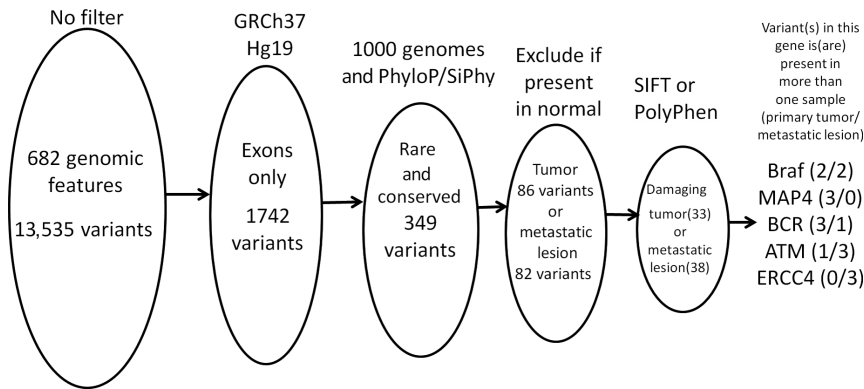
Figure 1

Schematic of the filtering criteria used on the genomic data to identify genes mutated in pathological samples. Five genes met an additional requirement that the gene had to be present in more than one sample.

were present in more than one sample (Fig. 2). Two variants and a deletion in *ATM* were distributed among five samples from four patients. A variant that results in an R3008C transition (CHR11:108236086) was found in a primary tumor and metastatic lesion, but not in matched normal tissue from the same patient. Variant CHR11:108236153 resulting in a G3030E was found in three samples from three different patients. Both *ATM* variants occur in the C-terminal FATC domain of the protein. We also identified a deletion in *ATM* resulting in a frameshift. Additionally, we identified two variants in the *ERCC4*/*XPF* tumor suppressor gene. Variant CHR16:14041714 was found in a metastatic lesion from HCC and was concurrent with *ATM* variant CHR11:108236086. The second variant as identified in two metastatic samples from two patients and is concurrent with *ATM* variant CHR11:108236153.

We compared our results in primary tumors with those of the TCGA project on differentiated PTC (Cancer Genome Atlas Research Network 2014). Approximately, 60% of TCGA samples have a *BRAF V600E* mutation and 13.5% *RAS* mutation. In contrast, our study found *BRAF V600E* in four samples and four samples with a *RAS* mutation among primary tumor and metastatic lesion samples. Although these differences may be due to the variety of histological types examined, a recent study focusing on poorly differentiated and anaplastic thyroid cancers (Landa *et al.* 2016) reported a similar rate of *BRAF V600E* (~33%) and *RAS* (~27%).

Interestingly, five samples in this study had variants in the *ATM* tumor suppressor. In TCGA, only 1.3% of the differentiated PTC harbored variants in this gene, and none of those were located in the FATC domain. *ATM* mutations were reported by Landa and coworkers in 9% of samples, although none were located in the FATC domain. In that study, four *ATM* samples were concurrent with *BRAF V600E*, and one sample had concurrent *ATM* and *NRAS Q61R* variants. Additional examination of publically available tissues (COSMIC) allowed us to identify 92 tumors and cell lines containing mutations of the *ATM* FATC domain. Five of those samples have
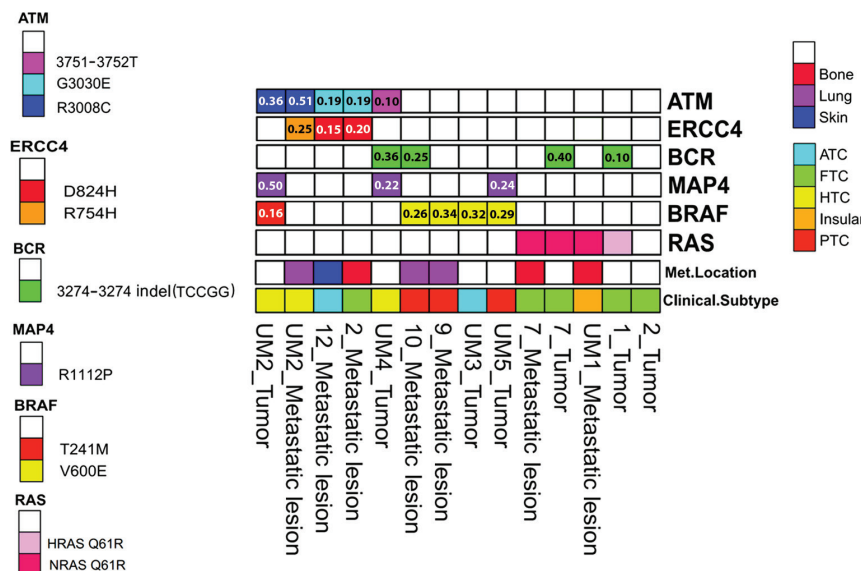
Figure 2

Candidate genes and variants found uniquely in more than one of 14 tumor tissue samples (primary or metastatic) vs normal tissues following filtering. One sample did not include a mutation in any of the analyzed genes.

concurrent BRAF mutations, but only two were *BRAF V600E*, one colon adenocarcinoma and one malignant melanoma. The FATC domain of ATM has been reported to be critical for telomere maintenance and effective DDR (Ogi *et al.* 2015). Additionally, the FATC domain has been implicated in *NOTCH1* regulation and leukemia cell survival (Vermezovic *et al.* 2015). Codon 3008 appears to be a mutation hotspot, and this variant has been shown to have a dominant negative effect *in vitro* (Meyn 1999). The relationship between *BRAF V600E* and *ATM* FATC domain variants may suggest redundant functions that would require experimental verification.

We also identified variants in a second gene involved in the DDR, *ERCC4/XPF*, exclusively in metastatic samples and only co-occuring with *ATM* FATC variants. Altered expression of *ERCC4* is common in colon cancer, where ~55% of cases have epigenetic repression of the gene (Facista *et al.* 2012). Although it is not clear that *ATM* and *ERCC4* cooperate in thyrocytes, such an interaction has been suggested in CLL cells in which co-occurrence of mutations in *ERCC4* and *ATM* predicted synthetic lethality to *ATR* kinase inhibitors (Kwok *et al.* 2016).

The PTC TCGA study identified DDR variants in ~6.5% of samples, and no case had mutations to multiple DDR components (Cancer Genome Atlas Research Network 2014). In contrast, Landa and coworkers reported in poorly differentiated tumors that 38% of samples contain a mutation in at least one DDR gene, most commonly *TP53*, and in ~6% of cases, there were mutations to multiple DDR genes (Landa *et al.* 2016). A recent study (Sohn *et al.* 2016) analyzed distant metastases and primary tumors from a group of well-differentiated PTC and FTCs using a targeted gene panel. That study found that *BRAF* and *RAS* mutations were largely conserved between primary tumors and distant metastases, but did not identify ATM variants or examine *ERCC4/XPF* as part of the panel. In this study, the *ATM* and *ERCC4* variants occurred in FTC, HCC or ATC tumors, whereas none were noted in PTC samples. Finally, due to its relevance in thyroid cancer, we examined primary tumor samples from four (10, 12, UM2 and UM4) patients with adequate residual DNA for *TERT* promoter mutations by Sanger sequencing. One C228T mutation was identified (UM4), which did not have a variant in *ERCC4* or a FATC domain variant in *ATM*. These data suggest that mutations in DDR-related genes may be enriched in poorly differentiated and non-PTC progressive metastatic thyroid cancers, although the small samples size of this study requires further work to test this hypothesis.

It is important to note that these findings are in a small number of samples from a diverse group of tumors and require confirmation to rule out false-positive results. Moreover, the use of a broader high-density sequencing platforms may lead to additional findings and functional studies are needed for variants of uncertain functional significance. We believe these descriptive data provide a scaffold for further studies to better define therapeutic targets for patients with progressive metastatic thyroid cancer.

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