The role of the inflammatory microenvironment in thyroid carcinogenesis

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Abstract

Immune responses against thyroid carcinomas have long been demonstrated and associations between inflammatory microenvironment and thyroid carcinomas repeatedly reported. This scenario has prompted scientists throughout the world to unveil how the inflammatory microenvironment is established in thyroid tumors and what is its influence on the outcome of patients with thyroid carcinoma. Many studies have reported the role of evasion from the immune system in tumor progression and reinforced the weakness of the innate immune response toward thyroid cancer spread in advanced stages. Translational studies have provided evidence that an increased density of tumor-associated macrophages in poorly differentiated thyroid carcinoma (DTC) is associated with an aggressive phenotype at diagnosis and decreased cancer-related survival, whereas well-DTC microenvironment enriched with macrophages is correlated with improved disease-free survival. It is possible that these different results are related to different microenvironments. Several studies have provided evidence that patients whose tumors are not infiltrated by lymphocytes present a high recurrence rate, suggesting that the presence of lymphocytes in the tumor microenvironment may favor the prognosis of patients with thyroid carcinoma. However, the effect of lymphocytes and other immune cells on patient outcome seems to result from complex interactions between the tumor and immune system, and the molecular pattern of cytokines and chemokines helps to explain the involvement of the immune system in thyroid tumor progression. The inflammatory microenvironment may help to characterize aggressive tumors and to identify patients who would benefit from a more invasive approach, probably sparing the vast majority of patients with an indolent disease from unnecessary procedures.

Introduction

Immune responses against differentiated thyroid carcinomas (DTCs) have long been demonstrated (Matsubayashi et al. 1995, Modi et al. 2003) and associations between inflammatory microenvironment and DTCs repeatedly reported (Yano et al. 2007, Muzza et al. 2010), evidenced by a mixture of lymphocytes and macrophages frequently found within and inside or surrounding primary thyroid tumors (Modi et al. 2003, French et al. 2010). These data support the hypothesis that immune responses may influence DTC progression.
In 2011, Hanahan & Weinberg (2011) stated that a cell acquires ten biological abilities to become a tumor and considered hallmarks of cancer the ability to sustain proliferative signaling, evade growth suppressors, avoid immune destruction, enable replicative immortality, promote inflammation, induce angiogenesis, activate invasion and metastasis, promote genome instability and mutation, resist cell death, and deregulate cellular energetics. Such abilities may appear for different reasons and can be instigated by diverse agents. In the case of thyroid carcinomas, risk factors may include ionizing radiations (Nikiforov 2010, Wartofsky 2010), chronic inflammatory conditions such as obesity (Zhao et al. 2012, Han et al. 2013), and ingestion of some nutrients such as iodine and others (Marcello et al. 2012). Individual genetically inherited profiles of susceptibility (Granja et al. 2004a,b, Bufalo et al. 2006, Guilhen et al. 2009, Xu et al. 2012) may explain different responses to both environmental and endogenous aggressions, as well as gender- and gene-associated alterations in the MAPK pathway and PI3K/AKT pathway or in cell-cycle genes, such as p53 (TP53), reflecting in clinical differences.

**Concepts of tumor immunology**

A close relationship between immune response and cancer was first proposed by Virchow in 1863 (Balkwill & Mantovani 2001). A few years later, Coley demonstrated that bacterial products were able to help inoperable cancer patients. The subsequent application of Bacillus Calmette-Guerin and other crude immunostimulants showed benefits that led to the regulatory approval of their use in the treatment of some solid tumors such as bladder cancer (Kirkwood et al. 2012). Thereafter, the capacity of the immune system to interfere with tumor progression has been proven by a series of clinical and epidemiological evidences. In fact, the success of Herceptin and Ipilimumab in treating breast and metastatic melanomas respectively has prompted scientists to go far beyond the frontiers in tumor immunology.

The field of tumor immunology is based on the fact that, during the tumorigenic process, malignant cells may express tumor-specific antigens and/or tumor-associated antigens. Tumor-associated antigens comprise gene products more frequently found in tumors, whereas tumor-specific antigens are gene products uniquely expressed in tumors (Lewis et al. 2003). These groups of antigens are target for immunotherapy, and new targets have actively been searched for all over the world. However, the absolute success of immunotherapy is impaired by tumor immune escape mechanisms.

Tumor cells are initially eliminated by the immune system before they become clinically detectable. This is then followed by an equilibrium phase, where a selection process for less immunogenic tumor variants takes place until tumor cells finally ‘escape’ from immune surveillance, as shown in Fig. 1 (Dunn et al. 2002).

One immune escape mechanism is active at the level of the tumor and can be ascribed to abnormalities in the expression of major histocompatibility complex (MHC) class I-restricted antigens, enabling tumors to take on a ‘stealth’ phenotype, hiding from immune cell detection (Poschke et al. 2011). The other mechanism results from the ability of the tumor to progress and ‘sabotage’ the host immune system. Certain tumors have the ability to use parts of the immune system to protect themselves against the host immune response. By doing so, tumors induce or recruit immune cells such as myeloid-derived suppressor cells (MDSCs) or regulatory T cells (Tregs). These cells physiologically serve as safeguards against overwhelming inflammation and lead to immune resolution (Poschke et al. 2011). The recruitment of these cells makes the microenvironment permissive for cell growth.
The immune system is a cornerstone determinant of the tumor microenvironment, as shown in Fig. 2. Recent molecular biology studies conducted in cancer patients have revealed that the inflammatory profile of the tumor microenvironment and, in particular, the acute inflammation of host tissues, may define patient prognosis (Hsu et al. 2010, Suzuki et al. 2011, Chew et al. 2012). Unraveling the biology and composition of the inflammatory microenvironment is crucial to understand the nuances of carcinogenesis, as well as to predict the clinical outcome of cancer patients. Herein, we describe the inflammatory microenvironment in thyroid carcinogenesis by dividing the issue functionally into innate immunity, adaptive immunity, and molecules of the immune system.

**Innate immunity**

Different from the adaptive immune response, innate immunity comprises mechanisms that protect the host in a nonspecific manner (Murphy 2011). Innate immunity is sustained by cells such as neutrophils, eosinophils, and mast cells, constituting the first line of defensive response. These cells are bone marrow-derived, tissue-dwelling granulocytes found transiently in the blood circulation en route to tissue inflammatory sites (Davoine et al. 2013). Special attention has been paid to the neutrophil-to-lymphocyte ratio (NLR).

NLR is derived from white blood cell differentiation counts (Chiang et al. 2012). Zahorec (2001) described the prognostic role of NLR in critically ill patients. Highest stress, marked neutrophilia, and lymphopenia were found in intensive care unit patients. Zahorec (2001) concluded that marked neutrophilia and lymphopenia are general innate immune responses to various stressful events. Marked neutrophilia and/or eosinophilia associated with neoplasia are relatively rare findings and are considered paraneoplastic manifestations only after excluding other
causes, such as infections, allergy, collagen disease, vascular diseases, and concomitant malignant hematopoietic diseases (Hardy & Balducci 1985). In addition, neutrophilia could represent a nonspecific response to cancer-related inflammation secondary to tissue destruction and cytokine release (Liu et al. 2013).

An association between high NLR and increased mortality or recurrence has been observed in various solid organ tumors (Hirashima et al. 1998, Walsh et al. 2005, Bhatti et al. 2010). Liu et al. (2013) investigated the total white blood cell and differential counts of 159 patients with DTC and 318 age- and sex-matched controls undergoing thyroidectomy for benign thyroid nodules. They found that cancer patients in the higher NLR tertile had a significantly larger tumor size and a high recurrence risk (Liu et al. 2013). There was no difference in NLR between patients having benign thyroid nodules and those having malignant thyroid nodules, suggesting that a high NLR is more important for tumor progression than for tumor initiation (Liu et al. 2013). In fact, experimental data indicate that activated neutrophils may directly and indirectly stimulate tumor growth (Fridlender et al. 2009). On the contrary, studying the NLRs of 26 patients with benign goiters, 31 patients with incidental papillary thyroid microcarcinoma, 26 patients preoperatively diagnosed with thyroid cancer, and 26 healthy controls, Seretis et al. (2013) found NLRs to be significantly elevated in patients with incidental papillary thyroid microcarcinoma and thyroid cancer, suggesting that the NLR should be considered an easily accessible biomarker for detecting incidental papillary thyroid microcarcinoma.

Since their discovery, a large number of studies have demonstrated natural killer (NK) cell-mediated lysis of different types of tumor cells in vitro, as well as NK cell-dependent elimination of many tumors in vivo (Ljunggren 2008). Phenotypically, NK cells are defined by the expression of CD16 and CD56 (NCAM1) surface markers (Cooper et al. 2001a). NK cells produce interferon γ (IFNγ) (IFNG), tumor necrosis factor α (TNFα (TNF)), TNFβ (LTA), and granulocyte-macrophage colony-stimulating factor (GMCSF (CSF2)) (Vivier et al. 2011). Gogali et al. investigated NK cell infiltration in the thyroid glands of 65 patients with PTC and 25 patients with thyroid nodular goiter (NG). They found a significantly increased number of NK cells in the PTC tissue samples than in the thyroid NG tissue samples. In addition, they demonstrated an inverse correlation between NK cell infiltration and tumor stage, with decreased NK cell infiltration in advanced stages of the disease (Gogali et al. 2012). This finding highlights the key role of evasion from the immune system in tumor progression and reinforces the weakness of the innate immune response toward cancer spread in advanced stages (Gogali et al. 2012).

However, does NK cell infiltration correspond to a homogeneous group of cells? The answer is probably no. There are two functionally different subsets of NK cells, CD56dim cells, which are cytotoxic; and CD56bright cells, which play an immunoregulatory role. These NK cell subsets present differences in their cytotoxic potential, cytokine production, and response to cytokine activation (Baume et al. 1992, Cooper et al. 2001b). The association of NK cells with cells that do not express MHC class I (e.g. tumor cells) makes the targets susceptible to NK cell-mediated lysis (Kare et al. 1986). Their ability to kill target cells, without prior sensitization, is regulated by the balance between stimulatory and inhibitory signals (Lanier 2008).

Liapi et al. (2013) investigated the distribution of CD3−CD16+CD56dim and CD3−CD16−CD56bright NK subpopulations in tissue and blood samples collected from patients with PTC and NG. Twenty-eight patients with PTC, 13 patients with NG, and 50 healthy donors were included in the study. The distribution of CD16−CD56bright and CD16+CD56dim NK cell subpopulations in the peripheral blood was similar in patients with PTC, patients with NG, and healthy donors. The number of CD16−CD56bright NK cells was increased in the tumor microenvironment of PTC tissue samples. Analysis of NK cell subpopulations in PTC tissue samples revealed that the number of CD16−CD56bright cells was higher than that of CD16+CD56dim cells. Comparison of NK cell subpopulations in the PTC and NG tissue samples revealed that the number of CD16+CD56dim cell was significantly higher in the NG tissue samples than in the PTC tissue samples. By contrast, CD16−CD56bright cells exhibited a higher infiltration in the PTC tissue samples than in the NG tissue samples, suggesting that an immunoregulatory pattern of NK cells is required for thyroid carcinogenesis (Liapi et al. 2013). Liapi et al. (2013) also found that the number of CD16+CD56dim NK cells was positively correlated with disease stage. The mean cell number in stage IVA was doubled when compared with that in stages I/II. The number of CD16−CD56bright NK cells was inversely correlated with disease stage and thyroid tissue infiltration, being greater in stages I/II. At first look, this result seems contradictory. However, it is well known that the tumor microenvironment of a stage IV tumor is quite different from that of an early-stage tumor (Cunha et al. 2012b). This fact should be considered along with the fact that NK cells migrating to lymphoid organs are CD56brighth NK cells and seem to be
immature in comparison with CD56dim NK cells (Lanier et al. 1986). Recent studies have confirmed that CD56bright NK cells are precursors of the CD56dim subset (Chan et al. 2007, Ouyang et al. 2007). The tumor microenvironment can be the site where immature CD56bright cells transform into mature cytotoxic NK cells, but this developmental program is not entirely fixed, and mature NK cells can be re-educated by local and environmental factors (Freud et al. 2006). This could probably explain the gradual increase or decrease in the percentages of NK cell subpopulations correlating with the tumor stage.

When tissue homeostasis is perturbed, macrophages and mast cells release soluble factors such as cytokines, chemokines, reactive oxygen species (ROS), and bioactive mediators such as histamine, which induces leukocyte migration and infiltration at the site of lesion, characterizing the inflammation (de Visser et al. 2006). When the resolution of this response fails, a disease can be initiated (Aggarwal et al. 2006, Medzhitov 2010). Active mast cells generate angiogenic factors such as vascular endothelial growth factor (VEGF), fibroblast growth factor, histamine, heparin, matrix metallopeptidase 9, and several proteases, in addition to leading to the release of more cytokines and chemokines (Ribatti et al. 2001). These data suggest that mast cells are critically important for tumor progression. In fact, Melillo et al. (2010) showed that human PTC tissues, but not normal thyroid tissues, present an intense mast cell infiltrate and that its intensity is positively correlated with the invasive behavior of thyroid carcinomas. They also found that thyroid carcinoma cell cultures are mast cell chemoattractants mediated by VEGFA. When injected in the tail vein of immunodeficient mice, mast cells were observed to infiltrate and that its intensity is positively correlated with the invasive behavior of thyroid carcinomas. An increased number of TAMs in PDTC was found to be associated with capsular invasion, extrathyroidal extension, and decreased cancer-related survival when compared with PDTC with a low density of TAMs, suggesting that TAMs could be associated with disease progression. In fact, the same group experimentally demonstrated that TAMs promote PTC progression (Ryder et al. 2013). Using BRAF-induced PTC mouse models, they observed that the conditional activation of BRAF in the thyroids of adult mice induces PTCs, which express macrophage chemoattractants and are densely populated with TAMs. When TAMs were depleted, PTC initiation was impaired. Phenotypic analysis demonstrated an increased expression of M2-related genes such as Ccr2, arginase 1, Ccl22, and Il10, whereas the expression of M1-specific markers (Il12, Il12b, and ROS (Ros1)) was not increased. Interestingly, selectively depleted TAMs during advanced stages of PTC induce tumor regression, and CSF1/CSF1R signaling can be pharmacologically targeted to impair PTC initiation (Ryder et al. 2013). These results suggest that TAMs may be rational therapeutic targets for patients with refractory advanced PTCs, particularly for those with PDTC and ATC.

Do macrophages play the same role in PDTC and well-DTC? They probably do not. We studied 398 patients with DTC and 132 with nonmalignant tissues (Cunha et al. 2012b). We found TAMs more frequently in more aggressive cases, with metastasis at diagnosis; however, paradoxically, macrophage infiltration was correlated with improved disease-free survival (Cunha et al. 2012b). As a protumorigenic role was proven previously, how to explain the apparent conflicting results? It is possible that
these different results are related to different micro-environments, as shown in Fig. 3 (Ryder et al. 2008). In fact, Fiumara et al. (1997) studied 121 well-differentiated PTC samples and found tumors with TAMs and in situ evidence of active neoplastic cell phagocytosis. Neoplastic cell phagocytosis by macrophages was positively correlated with the infiltration of both lymphocytes and dendritic cells (DCs), whereas it was negatively correlated with vascular invasion (Fiumara et al. 1997). They also found a trend of reduced risk of distant metastases at follow-up in cases with TAMs that, in addition, was associated with lymphocyte infiltration, confirming the complexity of the immunological host reaction to thyroid cancer phenomenon (Fiumara et al. 1997, Cunha et al. 2012b).

Adaptive immunity

Granulocytes and macrophages represent the effector mechanisms of innate immunity. Unfortunately, they cannot always eliminate infectious agents or even efficiently combat tumor development. Conversely, cells of the innate immune system play a crucial role in the initiation and subsequent direction of adaptive immune responses that are mediated by lymphocytes (Murphy 2011). The lymphocytes of the adaptive immune system have evolved to provide a more versatile defense system, which, in addition, provides increased protection against subsequent reinfection with the same pathogen (Murphy 2011).

How do tumor-infiltrating lymphocytes (TILs) interfere with tumor progression? CD4+ T cells are central to the successful orchestration of the immune response. Naive CD4+ T cells differentiate into one of at least four functionally distinct forms: Th1, Th2, Th17, or Tregs. In general, Tregs are identified as FoxP3+ lymphocytes and are thought to contribute to tumor-specific T-cell tolerance (Zhou & Levitsky 2007). The recent discovery of Th17 cells and their important role in host protection against infectious pathogens and in the pathogenesis of various inflammatory and autoimmune diseases have resulted in an explosion of immunological research. However, their role in human cancer is still under investigation (Su et al. 2010).

A mixture of lymphocytes are frequently found within and surrounding primary thyroid tumors (Modi et al. 2003, French et al. 2010). Villagelin et al. investigated 157 consecutive patients. They were classified by the degree of lymphocyte infiltration. Lymphocyte infiltration was classified as diffuse, peritumoral (only in or around the tumor), or absent. After a mean follow-up period of 8 years, they observed a significantly high recurrence in the absent lymphocyte infiltration group than in patients in the diffuse and peritumoral lymphocyte infiltration groups, suggesting that the presence of lymphocytes in the tumor microenvironment may favor the prognosis of patients with DTC (Villagelin et al. 2011), confirming previous reports (Matsubayashi et al. 1995, Gupta et al. 2001, Modi et al. 2003). Conversely, 100 PTC patients were analyzed for background thyroiditis and TILs by French et al. (2010). Patients with TILs exhibited higher disease stage and increased incidence of invasion and lymph node metastasis when compared with patients without lymphocytes or with background thyroiditis. The authors attributed these different results to the fact that concurrent thyroiditis would overshadow the real effect of lymphocytes on tumor progression. It is worth noting that considering only the presence and absence of TILs may obscure the functional and phenotypic diversity adjacent to different lymphocyte subsets.

To define the subset of lymphocytes found in association with PTC, French et al. (2010) analyzed primary tumors from ten patients with evident TILs by immunohistofluorescence. CD4+ T-cell frequency was found to be correlated with tumor size, Treg frequency with lymph node metastases, and CD8:Treg ratio inversely with tumor size. Our data demonstrated that the presence of concurrent chronic lymphocytic thyroiditis and infiltration of CD4+, CD8+, CD20+, Th17, and Treg cells are associated with favorable prognostic features in patients with DTC. Immune cells were found to infiltrate malignant tissues more often than benign lesions,
suggestions an immune reaction of the organism against transformed cells (Cunha et al. 2012b). Nevertheless, how
the immune system does act is not that simple. In fact, the
effect of the immune system on patient outcome seems
to result from complex interactions between tumor and
immune system cells.

Antitumor immune response is thought to be related
to tumor antigenicity. In fact, a correlation between protein
expression profile and immune cell infiltration in DTC
was observed (Bruland et al. 2009). Some researchers have
suggested that the expression of MUC1, NIS (SLC5A5),
ATM, PTEN, and CD56 might indicate tumor differen-
tiation and tumor progression, demonstrating tumor antigenicity (Larson et al. 2007, El Demellawy et al. 2008,
Morari et al. 2010). However, whether these markers are
really associated with DTC immune response is still not
clear. Aiming to examine the putative association between
tumor molecular profile and TIL, TAM, and MDSC patterns,
we investigated 398 patients whose tissue samples were
maintained in the tissue bank of the institution. Thyroid
carcinoma was diagnosed in 266 patients: 253 with PTCs
and 13 with follicular carcinomas. We also obtained 132
normal or benign thyroid tissue samples. Immune cell
infiltration was closely associated with the immuno-
histochemical profile of the DTC specimens examined,
including CD56, NIS, MUC1, PTEN, ATM, and B7H1
(CD274) (Cunha et al. 2012a).

The immune response rarely is enough to eliminate
tumors, and the reason for this is the existence of immune
escape mechanisms. One of these mechanisms is the
upregulation of B7H1 expression. B7H1 is a cell-surface
glycoprotein that may be involved in the regulation of
local inflammatory responses, and aberrant tumor
expression of B7H1 is thought to be associated with the
inhibition of the immune system, as shown in Fig. 4
(Hamanishi et al. 2007). The expression of B7H1 in tumor
cells has been reported to be associated with a poor
prognosis in some epithelial cancers (Thompson et al.
2006). We demonstrated that the expression of both B7H1
protein and B7H1 mRNA is upregulated in DTCs,
contrasting with the low levels displayed by benign tissues
(Cunha et al. 2013). This result suggests that tumor cells
may acquire B7H1 expression during the tumorigenic
process. We also observed an association between high
B7H1 mRNA levels and the presence of some features of
tumor aggressiveness, such as higher stages at presentation
and increased age at diagnosis, suggesting that high levels
of B7H1 expression may help identifying individuals who
need a more aggressive approach (Cunha et al. 2013). We
also studied the expression of B7H1 in 18 patients with
lymph node metastasis at diagnosis. Paired primary tumor
and lymph node metastatic tissue samples were obtained
from all the patients. We observed that the lymph node
metastatic tissue samples had lower levels of B7H1 than
the primary tumor tissue samples, indicating T-cell
exhaustion (Cunha et al. 2013). French et al. (2012) have
recently described high levels of IFN+ /CD8+ T cells in
12 metastatic lymph node tissue samples excised during
the initial surgery at patient presentation. The authors
found that proliferating lymphocytes were evident in
tumor-involved lymph node metastases that were
enriched with PD1 (ligand of B7H1) + lymphocytes
(French et al. 2012). They hypothesized that the presence
of metastases does not arrest and may, on the contrary,
promote an IFN+ response, suggesting the generation
of an antitumor response, which may impair tumor
evasion. In fact, this could explain the decrease in B7H1
expression that we observed in our matched metastases,
suggesting that T-cell exhaustion is a mechanism of
tumor progression.

The main players within the context of innate and
adaptive immunity are DCs, which induce, coordinate,
and regulate the system (Steinman 1991, Banchereau et
al. 2000). DCs are highly potent antigen-presenting cells
with the unique ability of taking up and processing antigens in
peripheral blood and tissues. They can migrate into the
draining lymph nodes, where they present antigens to
naïve T lymphocytes and thus induce a cellular immune
response involving CD4+ Th1 cells, cytotoxic CD8+ T cells,
and B cells (Schott 2006). As DCs can modulate the

Figure 4
Constitutive expression of B7H1 by exhausted T cells. Tumor cells may acquire B7H1. The interaction between tumor cells and T lymphocytes may induce an inhibition of the immune response mediated by B7H1 expression. B7H1 is associated with IL10 production, induction of Fas ligand expression, and high apoptosis rate.
whole immune repertoire, these cells are instrumental in the understanding of the inflammatory tumor microenvironment.

The importance of DCs in PTC was documented by an early study showing that patients with a dense infiltrate of S-100+ DCs in the tumor had a more favorable prognosis irrespective of other morphological and clinical features (Schroder et al. 1988). A series of 527 consecutive cases of thyroid carcinoma were investigated by Ugolini et al. aiming to detect DC infiltration. They found that the intratumoral inflammatory infiltrate (usually composed by mixtures of CD4+, CD8+, and CD20+ cells) was strongly reduced or absent in poorly differentiated and undifferentiated histotypes of thyroid carcinoma. DCs were most frequently detected in PTC cases (classical, follicular, and tall cell variants) and markedly reduced in poorly differentiated and undifferentiated tumors, pointing to the protective role of DCs and infiltrating lymphocytes against thyroid tumors (Ugolini et al. 2007).

In addition, Hilly et al. (2013) have recently demonstrated that DC density in papillary carcinoma is correlated with the concurrent thyroiditis grade and DC density in the surrounding areas of thyroiditis. These data suggest that the infiltration of DCs is part of a more complex series of outcome events that comprises lymphocyte infiltration and thyroiditis establishment.

Proietti et al. (2011) studied the infiltration of both immature CD1a+ DCs and mature DC-Lamp+ DCs in the intratumoral or peritumoral area, the differentiation of immature, but not mature, DCs was more frequently found in FVPTCs than in adenomas. Pusztaszeri et al. quantitatively assessed the presence of DCs that were positive for CD1a in the cytological samples of 31 histologically confirmed PTCs and in a control group of 29 benign thyroid nodules. CD1a-positive DCs were identified in 97% of the PTCs in thyroid cytology specimens. By contrast, only 31% of benign thyroid nodules had CD1a-positive DCs (Pusztaszeri et al. 2013). The significant presence of CD1a-positive DCs in PTCs suggests that these cells are recruited during malignant transformation.

Aiming to investigate this issue, Scarpino et al. established primary cultures of neoplastic thyroid cells and normal cells (from the tumor-free contralateral lobe of the same patients) obtained from eight thyroids removed surgically. Surprisingly, they found that normal thyroid cells and tumor cells were equally effective at releasing chemotactic activity for DCs in culture supernatants (Scarpino et al. 1999). This finding is in conflict with the observation that DCs are rare in normal thyroid tissues and raises the possibility that culture conditions may trigger stimulatory signals for normal thyroid cells. In fact, the same group had previously demonstrated that normal thyroid cells of primary cultures, but not normal thyroid cells of tissue sections, express high levels of MET protein and urokinase-type plasminogen activator receptor (uPAR) and both factors are engaged in DC recruitment (Rucho et al. 1996, Zanetti et al. 1998, Scarpino et al. 1999). MET protein is present in Hürthle cells of Hashimoto’s thyroiditis and hyperfunctioning thyrocytes of Graves’ disease (Rucho et al. 1996, Zanetti et al. 1998), indicating that the upregulation of MET receptor is an early event in thyroid cell alteration. This result corroborates the rare infiltration of DCs in benign thyroid lesions (Pusztaszeri et al. 2013).

### Immune system molecules in thyroid carcinogenesis

The immune system can produce or stimulate the production of many molecules in response to aggressive or nonproper features. These molecules are called cytokines and may be produced by signals of the innate or adaptive immune response for several types of cells. Cytokines influence the activation, growth, and differentiation of several different target cells, and more than 100 types of cytokines have been identified (Yadav et al. 2012). Table 1 summarizes the function of some cytokines. Failure to contain or impede injury can lead to persistent and excessive cytokine production, ultimately leading to harmful tissue destruction. Therefore, the balance of the host reactions to cellular stress, which will cause their production, might affect different stages of cancer (Dranoff 2004).

### Interferon γ

IFNγ is a cytokine that mediates pleiotropic effects on the innate and adaptive responses to infection, and its deficiency or deficiency of its receptor has been reported to be related to the development of more tumors in mice exposed to chemical carcinogens (Bach et al. 1997, Smyth et al. 2000). IFNγ has long been reported to be associated with cytostatic/cytotoxic and antitumor functions (Brown et al. 1987). Kaplan et al. (1998) reported that approximately one-third of melanoma and lung adenocarcinoma cell lines have inactivating mutations in IFNγ pathway components, suggesting that tumor insensitivity to this cytokine could be an evasion mechanism. Recombinant IFNγ treatment was subsequently tested to...
Table 1  Summary of the effects of some cytokines and the cells in which they are produced

<table>
<thead>
<tr>
<th>Molecule</th>
<th>Producer</th>
<th>Effect</th>
<th>References</th>
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| IFNγ     | Activated macrophages | Cytostatic/cytotoxic  
Cytoproliferative  
Stimulation of the secretion of  
Th1 cytokines | Brown et al. (1987) and Rotondi et al. (2013) |
|          | Tumor cells | Apoptosis  
Necrosis  
Angiogenesis  
Immune cell activation  
Differentiation  
Cell migration  
| TNF      | Activated macrophages | Stimulation of the secretion of  
Th1 cytokines | Beutler et al. (1985), Wajant (2009) and Lippitz (2013) |
|          | CD4+ lymphocytes | Necrosis  
Angiogenesis  
Immune cell activation  
Differentiation  
Cell migration | Beutler et al. (1985), Wajant (2009) and Lippitz (2013) |
|          | NK cells | Necrosis  
Angiogenesis  
Immune cell activation  
Differentiation  
Cell migration | Beutler et al. (1985), Wajant (2009) and Lippitz (2013) |
|          | Tumor cells | Necrosis  
Angiogenesis  
Immune cell activation  
Differentiation  
Cell migration | Beutler et al. (1985), Wajant (2009) and Lippitz (2013) |
| TGFβ     | Myeloid cells | Epithelial-to-mesenchymal transition  
Metastasis  
Immune tolerance  
Cell survival | Arsura et al. (2003), Bierie & Moses (2006) and Fabregat et al. (2013) |
|          | T lymphocytes | Metastasis  
Immune tolerance | Arsura et al. (2003), Bierie & Moses (2006) and Fabregat et al. (2013) |
|          | Tumor cells | Metastasis  
Immune tolerance | Arsura et al. (2003), Bierie & Moses (2006) and Fabregat et al. (2013) |
| IL1      | Leukocytes | Proliferation  
Differentiation  
Proangiogenic  
Prometastatic  
Production of inflammatory molecules  
Stimulation of antibody synthesis  
Proangiogenic  
Prometastatic  
Production of inflammatory molecules  
Stimulation of antibody synthesis | Waldmann (2006) |
|          | Tumor cells | Proliferation  
Differentiation  
Proangiogenic  
Prometastatic  
Production of inflammatory molecules  
Stimulation of antibody synthesis | Waldmann (2006) |
| IL2      | CD4+ lymphocytes | Proliferation of NK cells  
Differentiation of NK cells | Waldmann (2006) |
|          | CD8+ lymphocytes | Proliferation of NK cells  
Differentiation of NK cells | Waldmann (2006) |
|          | Activated DCs | Proliferation of NK cells  
Differentiation of NK cells | Waldmann (2006) |
|          | Activated NK cells | Proliferation of NK cells  
Differentiation of NK cells | Waldmann (2006) |
| IL4      | Th2 cells | Response against helminths  
Suppression of Th1 cells  
Induction of IgE class switching in B cells  
Expression of class II MHC  
Expression of B-cell receptors  
Prolongation of the lifespans of T and B cells  
Mediation of tissue adhesion and inflammation | Howard et al. (1982) and Akdis et al. (2011) |
|          | Basophils | Leukocyte trafficking  
Leukocyte activation  
Induction of acute-phase protein production  
T-cell proliferation  
B-cell differentiation  
B-cell survival | Hurano et al. (1985) and Hurst et al. (2001) |
|          | Mast cells | Leukocyte trafficking  
Leukocyte activation  
Induction of acute-phase protein production  
T-cell proliferation  
B-cell differentiation  
B-cell survival | Hurano et al. (1985) and Hurst et al. (2001) |
|          | Eosinophils | Leukocyte trafficking  
Leukocyte activation  
Induction of acute-phase protein production  
T-cell proliferation  
B-cell differentiation  
B-cell survival | Hurano et al. (1985) and Hurst et al. (2001) |
|          | Tumor cells | Leukocyte trafficking  
Leukocyte activation  
Induction of acute-phase protein production  
T-cell proliferation  
B-cell differentiation  
B-cell survival | Hurano et al. (1985) and Hurst et al. (2001) |
| IL6      | Endothelial cells | Leukocyte trafficking  
Leukocyte activation  
Induction of acute-phase protein production  
T-cell proliferation  
B-cell differentiation  
B-cell survival | Hurano et al. (1985) and Hurst et al. (2001) |
|          | Fibroblasts | Leukocyte trafficking  
Leukocyte activation  
Induction of acute-phase protein production  
T-cell proliferation  
B-cell differentiation  
B-cell survival | Hurano et al. (1985) and Hurst et al. (2001) |
|          | Monocytes | Leukocyte trafficking  
Leukocyte activation  
Induction of acute-phase protein production  
T-cell proliferation  
B-cell differentiation  
B-cell survival | Hurano et al. (1985) and Hurst et al. (2001) |
|          | Macrophages | Leukocyte trafficking  
Leukocyte activation  
Induction of acute-phase protein production  
T-cell proliferation  
B-cell differentiation  
B-cell survival | Hurano et al. (1985) and Hurst et al. (2001) |
| IL10     | Monocytes | Immune resolution  
Immune tolerance  
Impairment of T-cell activation  
Survival, proliferation, and differentiation of human B cells | Akdis et al. (2011) |
|          | T cells (mainly Th2 cells) | Immune resolution  
Immune tolerance  
Impairment of T-cell activation  
Survival, proliferation, and differentiation of human B cells | Akdis et al. (2011) |
|          | B cells | Immune resolution  
Immune tolerance  
Impairment of T-cell activation  
Survival, proliferation, and differentiation of human B cells | Akdis et al. (2011) |
|          | NK cells | Immune resolution  
Immune tolerance  
Impairment of T-cell activation  
Survival, proliferation, and differentiation of human B cells | Akdis et al. (2011) |
|          | Activated DCs | Immune resolution  
Immune tolerance  
Impairment of T-cell activation  
Survival, proliferation, and differentiation of human B cells | Akdis et al. (2011) |
|          | DCs | Immune resolution  
Immune tolerance  
Impairment of T-cell activation  
Survival, proliferation, and differentiation of human B cells | Akdis et al. (2011) |
|          | Mast cells | Immune resolution  
Immune tolerance  
Impairment of T-cell activation  
Survival, proliferation, and differentiation of human B cells | Akdis et al. (2011) |
|          | Tumor cells | Immune resolution  
Immune tolerance  
Impairment of T-cell activation  
Survival, proliferation, and differentiation of human B cells | Akdis et al. (2011) |
| Leptin   | Adipocytes | Activation of monocytes and macrophages  
Angiogenesis  
Suppression of the production of anti-inflammatory cytokines | Kwon & Pessin (2013) |
| Resistin | Adipocytes | Production of IL6 and TNFα | Bokarewa et al. (2005) |
| Adiponectin | Adipocytes | Insulin sensitivity  
Inhibition of LPS-induced TNFα production in macrophages  
Differentiation of M2 macrophages  
Stimulation of the production of anti-inflammatory IL10  
Modulation of T-cell activation and inflammatory function of NK cells | Yokota et al. (2000), Kumada et al. (2004), Takemura et al. (2007) and Li et al. (2009) |
Treat a variety of tumors: chronic myelogenous leukemia, bladder carcinoma, colorectal cancer, melanoma, ovarian cancer, and adult T-cell leukemia, but the results were considerably heterogeneous (Kurzrock et al. 1987, Kloke et al. 1992, Miller et al. 2009), suggesting that IFN\(\gamma\) has, in fact, two faces: it can have cytostatic/cytotoxic and cytoproliferative effects, depending on the context, similar to transforming growth factor \(\beta\) (TGF\(\beta\)) and TNF, which also display this kind of dual contrasting behaviors (Roberts & Wakefield 2003, Wajant 2009). IFN\(\gamma\) studies in thyroid tissues have been carried out more frequently in autoimmune diseases, such as Hashimoto’s thyroiditis. Some authors have reported that transgenic mice expressing interferon in the thyroid gland develop numerous features of Hashimoto’s thyroiditis, including goiter, disrupted thyroid architecture, long-lasting hypothyroidism, and Hürthle cell metaplasia, indicating that IFN\(\gamma\) can transform a thyrocyte into a Hürthle cell by inducing the expression of an immunoproteasome component called LMP2 (PSMB6; Caturegli et al. 2000, Kimura et al. 2005, 2009). In a recent study, Rotondi et al. (2013) have shown that IFN\(\gamma\) stimulates the secretion of CXCL10, a situation associated with Th1 T-cell infiltration typical of autoimmune thyroid disease (AITD), which is closely related to thyroid tumors, leading, in most cases, to a better prognosis of patients (Cunha & Ward 2011, 2012). Conversely, the administration of TNF\(\alpha\) has been shown to lead to high levels of CXCL8 (IL8), a chemokine that is not primarily involved in thyroid autoimmunity, but that might induce tumor-related inflammatory infiltrating lymphocytes in thyroid diseases.

**Tumor necrosis factor**

TNF has a central role in immune homeostasis, inflammation, and host defense. Depending on the cellular context, it can induce diverse processes such as apoptosis, necrosis, angiogenesis, and immune cell activation, differentiation, and migration (Wajant 2009). TNF can also modulate cachexia, mediating cancer-associated fatigue and muscle wasting by its proinflammatory properties (Beutler et al. 1985). TNF\(\alpha\) and interleukin 1 (IL1) are described as acute-response cytokines. They activate vascular endothelial cells and induce endothelial cells to express adhesion molecules for neutrophils, monocytes, and lymphocytes (Lippitz 2013). Although TNF has a strong role in apoptosis and necrosis, systemic clinical tests with this factor have failed due to the severe inflammatory side effects associated with systemic TNF receptor activation (Wajant 2009). Nevertheless, safe local administration of TNF in combination with melphalan by isolated limb perfusion was tested as a treatment option for locally advanced soft limb sarcomas, and there are ongoing clinical studies aiming to identify antibodies able to restrict TNF activity to the tumor area or to inhibit the therapy-limiting side effects (van Horssen et al. 2006). Increased serum concentrations of TNF\(\alpha\) were described in eight tumors: non-small-cell lung carcinoma (NSCLC), breast cancer, colorectal cancer, prostate cancer, chronic lymphocytic leukemia, malignant melanoma, non-Hodgkin lymphoma, and gastric cancer (Lippitz 2013). These studies provide evidences that the production of TNF\(\alpha\) in the tumor microenvironment can lead to direct DNA damage; has an apoptotic or antiapoptotic role and, depending on downstream signaling, mitogenic activity; can mediate tumor cell/stroma relationships; and can induce other cytokines and chemokines that promote tumor development (Vendramini-Costa & Carvalho 2012). Zhang et al. (2013) have recently demonstrated that the administration of thyroid-stimulating hormone (TSH) can increase TNF\(\alpha\) expression, which has already been reported to be related to the chemokine profile of tumor inflammation, possibly indicating that TNF\(\alpha\) might play an indirect role in thyroid tumors by the inflammation that it stimulates (Rotondi et al. 2013). In Graves’ disease, inflammatory mediators, such as ILs and TNF\(\alpha\), stimulate the production of external thyroid-stimulating antibodies that bind to the TSH receptor (Baur et al. 2000, Davis 2008).

**Transforming growth factor \(\beta\)**

TGF\(\beta\) is a member of the growth factor TGF\(\beta\) superfamily, including bone morphogenetic protein, activins, inhibins, and anti-Müllerian hormone, among others, produced by cancer cells, myeloid cells, and T lymphocytes. TGF\(\beta\) is best known for its antiproliferative effects, and its signaling is an important regulator of the epithelial-to-mesenchymal transition and metastasis (Bierie & Moses 2006). However, tumor cells develop mechanisms to overcome TGF\(\beta\)-induced suppressor effects. Once this occurs, cells may respond to this cytokine inducing other effects that contribute to tumor progression, such as immune tolerance that shields the tumor from immune surveillance (Vendramini-Costa & Carvalho 2012, Fabregat et al. 2013). Numerous studies have established that deregulated activation of the NF-κB and PI3K/AKT pathways by TGF\(\beta\) promotes cancer cell survival (Arsura et al. 2003, Lin et al. 2007). Interestingly, TGF\(\beta\) routinely suppresses the activation of NF-κB in normal epithelial...
cells by inducing their expression of IκBα (NFκBIA; Nagumo et al. 2005, Sanchez-Capelo 2005). However, this event is inactivated during oncogenic progression, and TGFβ stimulation of late-stage cancers of the breast, prostate, and liver has been proven to result in NF-κB activation, leading to the induction of prosurvival and tumorigenic gene expression profiles (Arsura et al. 2003). Both TGFβ mRNA and TGFβ protein are expressed in thyroid cells. There is evidence that iodine and TSH are regulators of this production. Experimental studies have shown that iodine excess leads to an increased mRNA expression of TGFβ, stimulating protein synthesis (Yuasa et al. 1992, Cowin & Bidey 1995). In addition, studies in rats treated with TRH showed that during the formation of goiter, there is an increase in Tgfβ gene expression (Morosini et al. 1996). Kimura et al. (1999) proved that goiter and DTC tissues exhibit a higher expression than normal tissues. In a more recent study, Eloy et al. (2012) have proposed a new division of malignant tumors, classifying them into well-circumscribed carcinomas (WCPTCs), ones lacking infiltrative features and lymph node metastasis, and poorly circumscribed carcinomas, ones having these characteristics. This allowed the authors to observe that TGFβ was less expressed in the center of WCPTCs and that this expression increased as they became invasive forms, also associated with SMAD4 and SMAD7 proteins, probably increasing the ability of these tumors to progress and invade. These findings provided new perspectives for the understanding of TGFβ mechanisms for tumor progression, as not only the classically studied pathways, such as PI3K/AKT and MAPK, appear to be involved in malignant TGFβ transformation, but also SMAD molecules might play a role in this event.

Interleukins

A large group of secreted proteins that bind to their specific receptors and play a role in the communication among leukocytes are called ILs, and they are divided into families based on sequence homology and receptor chain similarities or functional properties (Akdis 2011). Some of the immune system molecules, such as CD4+ Th cells, are classified according to the cytokine profile of the microenvironment where they are present, which is ultimately regulated by the expression of other cytokines, showing a dependence of some ILs on other cytokines to be produced (Fan et al. 1998, Akdis 2011). Such a profile might define the specific properties of the cells and their role in immune response to nonproper agents.

IL1 is a proinflammatory cytokine that mediates many inflammatory diseases and interferes in cell proliferation, differentiation, and function in innate and adaptive immunity (Akdis 2011). IL1 can be produced by neoplastic cells or by the microenvironment where it is present, stimulating the production of proangiogenic and prometastatic mediators (Lewis et al. 2006). As it has a role in proliferation and can also lead to the production of ROS and RNS, IL1 can favor the accumulation of mutations, increase the invasiveness of tumor cells that are existing already, activate the angiogenic machinery, and induce the production of inflammatory molecules, contributing to the spread of tumor cells (Apte et al. 2006). Thus, as expected, high levels of IL1 would be associated with a worse prognosis. In fact, the expression of IL1 has been shown to be upregulated in melanomas and colon, lung, head-and-neck, and gastric cancers (Perrier et al. 2009).

With regard to thyroid tissues, Rebuffat et al. (2013) have recently demonstrated that IL1β (IL1B) modifies thyroid epithelial tightness by altering the expression and localization of junction proteins in Graves’ and Hashimoto’s diseases, suggesting that IL1β could play a role in the pathogenesis of thyroid autoimmunity. Alterations in serum concentrations of IL1β have been suggested to differentiate PTCs from atrophic thyroiditis (Kammoun-Krichen et al. 2012). The inheritance of the polymorphism rs2192752 in IL1 receptor (IL1R) increases the risk of PTC, indicating an association of this IL with DTC (Park et al. 2012).

Several studies have also been conducted to test the efficacy of IL2 in stimulating the immune system of cancer patients, as the main functions of IL2 are as follows: regulating the growth of B cells, stimulating the synthesis of antibodies, and increasing the cytolytic capacity of cells, through the increase in NK cell proliferation (Waldmann 2006). As a matter of fact, recombinant human IL2 is used in immunotherapy for advanced renal cancers and melanomas and for treating AIDS associated with HIV (Akdis et al. 2011, Liang et al. 2012). The administration of high doses of IL2 is associated with a better response rate in the case of melanomas and renal cancers, leading to improvements in disease-free survival, although the side effects of IL2 administration are remarkably hard on most of the patients (Clement & McDermott 2009, Halama et al. 2010).

IL4 is produced by Th2 cells, basophils, mast cells, and eosinophils and, similar to most of the cytokines, is able to regulate other immune molecules (Howard et al. 1982). IL4 is one of the most important cytokines for the differentiation of CD4+ cells into Th1 and Th2 cells, essential
effectors of immune responses (Akdis et al. 2011). Besides Th1 and Th2 cell production, IL4 also affects the phenotypes of B and T cells, leading to prolonged cell lifespans, which will affect tissue adhesion and inflammation (Akdis et al. 2011). Studies have long shown that IL4 has an inhibitory effect on the growth of human melanomas, renal cell carcinomas, and gastric cells (Hoon et al. 1991). On the contrary, IL4 and IL10 have been reported to exert a stimulatory effect on the growth of thyroid cancer cells. The production of IL4 and IL10 has been shown to be related to the promotion of thyroid tumor cell progression through the downregulation of BCL2 and BCL-XL (BCL2L1), leading to the death of a tumor cell extinction when they neutralized these cytokines, suggesting that concomitant Graves’ disease and PTC that exhibited IL4 and IL10 expression were more likely to display apoptosis resistance, probably potentiating anti-apoptotic factors, such as insulin-like growth factors. In the same way, Todaro et al. demonstrated that IL4 and IL10 upregulate cFLIP (CFLAR) and PED/PEA15, neutralizing the interaction of CD95L with CD95, which ultimately results in thyroid tumor cell survival and proliferation. The authors observed thyroid tumor cell extinction when they neutralized these cytokines, suggesting that they could be used as therapeutic targets (Todaro et al. 2006). In a recent study, Lee (2012) has described the induction of DCs from peripheral blood mononuclear cells purified from patients with thyroid cancer by culturing them in the presence of FL, GMCSF, IL4, and TNFα. The authors provided morphological evidence that the co-culture of T cells/cancer tissues activated T cells and differentiated cytotoxic T lymphocytes. These cytotoxic T lymphocytes adhered to cancer tissues and lysed cancer tissues, suggesting that DCs could be used as potential vaccines in immunotherapy, resulting in minimum residual disease after the conventional treatments applied to cancer (Lee 2012).

IL6 is involved in both innate and acquired immunity and its production is affected by stimuli from other cytokines (IL1, IL17, and TNFα; Akdis 2011). In innate immunity, IL6 is considered a regulator of acute-phase responses, through the activation of leucocytes and also the stimuli for the expression of acute-phase proteins (Hurst et al. 2001). IL6 also participates in acquired immunity, promoting B-cell differentiation and survival and plasma-cell production of antibodies (Hirano et al. 1985, Akdis 2011). There are strong positive associations between serum IL6 concentrations and tumor size, tumor stage, and disease progression in patients with gastric cancer, colorectal cancer, bone sarcoma, breast cancer, hepatocellular cancer, nasopharyngeal cancer, renal cell cancer, lung cancer, and melanoma (Niitsu et al. 2002, Chow et al. 2003, Nikiteas et al. 2005, Tas et al. 2005, Ahmed et al. 2006, Hsia et al. 2006, Egler et al. 2008, Ikeguchi et al. 2009). Although there are several evidences that IL6 is closely related to AITDs, its relationship with thyroid cancers has not been fully investigated (Gillespie et al. 2012, Popova et al. 2012, Beumer et al. 2013).

IL10 is an anti-inflammatory factor and an important regulator of several aspects of immune responses (Akdis 2011). It is very important in the microenvironment where it is located, since it inhibits the expression of many proinflammatory cytokines, chemokines, and chemokine receptors, thus influencing the production and effects of a large number of molecules, indirectly affecting T-cell activation (Akdis 2011). Circulating concentrations of IL10 were investigated in different cancer types and were found to be associated with adverse disease stage or with negative prognosis in bone sarcoma, diffuse large B-cell lymphoma, gastric cancer, colon cancer, Hodgkin’s lymphoma, hepatocellular cancer, melanoma, renal cell cancer, NSCLC, and pancreatic cancer (Sato et al. 1996, Onishi et al. 1999, Suzuki et al. 2001, Chen et al. 2013). Our group showed that the inheritance of a G allele at IL10 – 1082A/G polymorphism may favor concurrent thyroid autoimmunity in DTC patients, and this autoimmunity may favor a better prognosis of these patients (Cunha et al. 2011). In addition, Yu et al. (2013) demonstrated that the concentration of IL10 was significantly higher in patients with PTCs associated with multinodular goiter (MNG) than in patients with MNG alone, suggesting that cancer patients would have a specific type of Treg that affects antitumor responses and may facilitate disease progression and worse prognosis.

Adipokines

Adipokines or adipocytokines are cytokines produced by the adipose tissue. They have different functions, such as regulation of appetite and energy balance, immunity, insulin sensitivity, angiogenesis, inflammation and acute-phase response, blood pressure, and lipid metabolism (Kwon & Pessin 2013). Many current researches involve adipokines such as leptin, adiponectin, and resistin, considering that these adipokines have a direct relationship with the adipose tissue and are directly involved in changes in the immune response and insulin resistance, which can be a risk factor of several diseases associated
with obesity, such as diabetes. Our group has obtained preliminary data suggesting a relationship between obesity and DTC development (Marcello et al. 2012), but the molecular mechanisms involved in this relationship are not yet very well understood, although recent literature has been pointing to insulin resistance and potentially adipokines as probable connectors.

The structure of leptin is similar to that of IL2, IL6, and granulocyte colony-stimulating factor, and similar to other cytokines, it can activate monocytes and macrophages, stimulate VEGF and angiogenesis, and suppress the production of anti-inflammatory cytokines (Kwon & Pessin 2013). The induction of inflammatory responses by leptin involves its receptor b (LepRb), JAK2, and STAT3 signaling pathway (Kwon & Pessin 2013), and all these molecules ultimately have an effect on the PI3K/AKT and MAPK signaling pathways. Thus, leptin interacts with several factors that participate in the main signaling pathways of DTCs and could represent one of the links between obesity and DTC. The expression of leptin and its receptor, OBR (LEPR), has already been reported to be associated with a high risk of lymph node metastases, worsening the prognosis of patients, suggesting that the participation of this adipokine in DTCs might be important for their progression (Cheng et al. 2010).

Resistin is primarily related to insulin resistance by the activation of SOCS3 and the suppression of insulin-mediated signaling in rat adipocytes (Steppan et al. 2005), but in humans this association is not always true. This might be because resistin is produced not only by adipocytes (as in rodents) but also by monocytes and macrophages in humans (Kwon & Pessin 2013). In macrophages, the expression of resistin is induced by inflammatory cytokines such as IL1β, IL6, TNFα, and lipopolysaccharide (LPS). In human peripheral mononuclear cells, resistin stimulates the production of IL6 and TNFα through the NF-κB signaling pathway (Bokarewa et al. 2005). Similar to leptin, resistin also binds to TLR4, activating JNK and p38 MAPK to induce insulin resistance (Benomar et al. 2013). The expression of resistin has been reported to be associated with the increased proliferation of prostate cancer cells by the stimulation of the AKT pathway, a widely studied pathway in follicular thyroid tumors (Kim et al. 2011). There are also reports of the influence of resistin on breast cancer cells (Kang et al. 2007, Sun et al. 2010). As for other adipokines, there are evidences that the expression of resistin can be modulated by the presence of polymorphisms in its gene and gene receptor (Chen et al. 2010). Eke Koyuncu et al. (2013) have recently demonstrated significant correlations between resistin and TSH levels in hyperthyroid individuals, suggesting that the levels of this cytokine are directly related to thyroid dysfunction.

Adiponectin is highly expressed by adipocytes with potent anti-inflammatory properties and its expression can be suppressed by proinflammatory factors such as TNFα, IL6, ROS, and hypoxia (Li et al. 2009). The administration of adiponectin or its overexpression in transgenic mice results in improved insulin sensitivity, inhibits LPS-induced TNFα production in macrophages, promotes the differentiation of anti-inflammatory M2 macrophages and phagocytosis to remove apoptotic cells, stimulates the production of anti-inflammatory IL10, and also modulates T-cell activation and inflammatory function of NK cells (Yokota et al. 2000, Kumada et al. 2004, Takemura et al. 2007). Recently, Mitsiades et al. (2011) have demonstrated that serum adiponectin concentrations are inversely correlated with DTC, exerting a protective effect against the development of this cancer. Furthermore, the authors demonstrated that thyroid tissues express the receptors ADIPOR1 and ADIPOR2, which facilitate the entrance and functioning of adiponectin, suggesting a relationship between this cytokine and thyroid tissues.

In summary, DTCs are enriched with multiple inflammatory cells and both innate and adaptive immune responses seem to be engaged in thyroid carcinogenesis. In addition, the molecular pattern of cytokines and chemokines helps to explain the involvement of the immune system in tumor progression. The first attempt to use the immune system as a tool for treating patients with thyroid cancer was made in 1975. Active immunotherapy was applied to three patients. Two of them, who were in the terminal stage of the disease, could not develop generalized cell-mediated immunity. One patient developed in vitro evidence of cell-mediated immunity against cancer tissue antigens, associated with a decrease in tumor size (Amino et al. 1975). Unfortunately, more than 30 years later, no relevant successful result has been obtained for thyroid immunotherapy. On the other hand, the inflammatory microenvironment may help to identify more aggressive tumors and patients who would benefit from a more invasive approach, probably sparing the vast majority of patients with an indolent disease from unnecessary procedures.

Declaration of interest
The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the review reported.


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